

Synthesis of deacetyl-1,10-didehydrosalvinorin G

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Abstract

To unambiguously confirm the actual product in autoxidation of salvinorin A under basic conditions, deacetyl-1,10-didehydrosalvinorin G was synthesized from salvinorin C via intermediate salvinorin H. Furthermore, oxidation of salvinorin D with manganese dioxide gave salvinorin G in good yield.

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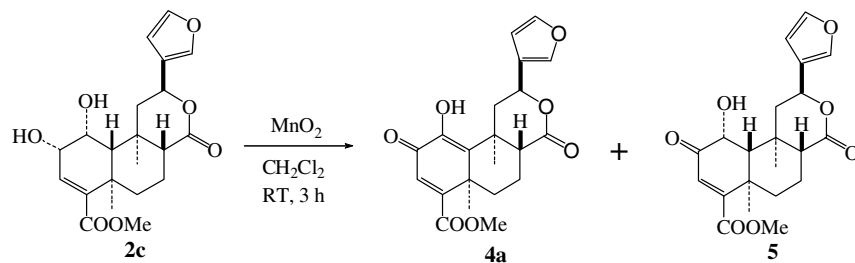
Keywords: Salvinorin A; Oxidation; Manganese dioxide; Synthesis

Salvia divinorum, a Mexican medicinal plant, has been used traditionally for its psychoactive (hallucinogenic) effects in divination rites.¹ Previous phytochemical studies have resulted in the isolation of 34 compounds, including salvinorins A (**1a**), C (**2a**), D (**2b**), G (**3**), and H (**2c**).^{2–9} Of those compounds, **1a** was identified as a potent and selective kappa opioid receptor (KOR) agonist.^{10,11} Because of the unique non-nitrogenous structure and potent binding activities to KOR, much effort has been directed toward a better understanding of structure–activity relationships (SAR) of **1a**.^{12–26} Salvinorin derivatives readily underwent epimerization at C-8 under basic conditions.^{3,4,13–17} Surprisingly, treatment of **1a** and its derivative with strong bases, such as Ba(OH)₂,¹⁵ KOH,^{18,26} and NaOH,²⁵ yielded corresponding natural salvinorin analogs, and no epimerization at C-8 was observed. It was reported that treatment of **1a** with KOH in methanol produced deacetyl-1,10-didehydrosalvinorin G (**4a**).¹⁸ Recently, we revised the structure of **4a** to its 8-epimer (**4b**) based on comparison of ¹H and ¹³C NMR data with those of **1a** and **1b**, NOESY data, and chemical conversion.²⁷ To unambiguously confirm the actual product in autoxidation of **1a** under harsh basic conditions,¹⁸ it is necessary to syn-

thesize the natural salvinorin derivative **4a** and to complete its NMR data. In this Letter, we report the synthesis of **4a** from **2a** via intermediate **2c**, and chemical conversion of **3** from **2b**.

Following the published procedure,¹⁸ the diol **2c** was prepared by deacetylation of **2a**. Subsequent oxidation of **2c** with manganese dioxide (Scheme 1) yielded **4a** and deacetylsalvinorin G (**5**).²⁸ Using 2D NMR techniques, including COSY, NOESY, HMQC, and HMBC, permitted the full assignment of all ¹H and ¹³C NMR chemical shifts of **4a** (Tables 1 and 2), and the key ¹³C–¹H correlations in the HMBC spectrum of **4a** are shown in Figure 1. In the ¹H NMR spectrum of **4a**, the C-8-H shifted much upper field to δ 2.42, and the coupling constants (dd, $J = 12.6$ and 3.3 Hz) of H-8 are more suitable for axial orientation than those (δ 2.99, dd, $J = 9.6$ and 5.1 Hz) of **4b** (Table 1). The C-12-H of **4a** shifted low-field slightly compared with that of **4b** (Table 1), and the H-12 of **4a** showed the J values (10.5 and 6.6 Hz) for axial orientation. In addition, the H-11 α (δ 3.76) of **4a** shifted much lower field compared with that of **4b** (δ 3.11), and the chemical shift changes are consistent with those of **2a** (δ 2.49)⁴ and its 8-epimer (δ 2.14).²⁹ Comparison of the ¹³C resonances of C-6, C-8, C-12, C-13, C-17, C-19, and C-20 (Table 2) of **4a** and **4b** also confirms that H-8 in **4a** is the β configuration. In the NOESY spectrum of **4a**, H-12 (δ 5.62) showed cross peaks to H-11 α (δ 3.76) and H-20 (δ 1.54), while H-19 (δ 1.76)

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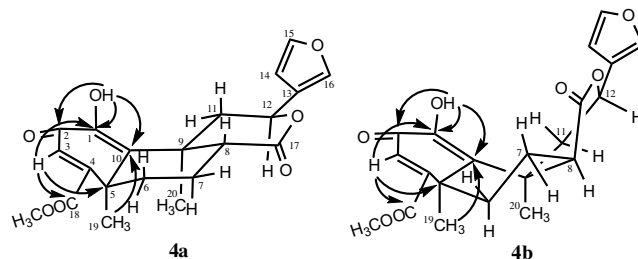


Scheme 1.

Table 1

¹H NMR data (300 MHz, CDCl₃) for **4a** and **4b** [δ (ppm), m, *J* (Hz)]

Proton	4a	4b
3	6.88 s	7.00 s
6 α	2.51 dt (13.2, 3.3)	2.54 dt (13.8, 7.5)
6 β	1.46 td (13.5, 3.9)	1.67–1.77 m
7 α	1.94 ddt (14.4, 3.3, 13.5)	2.24 ddd (14.4, 7.8, 5.1)
7 β	2.27 dq (14.7, 3.6)	1.98 ddd (14.1, 9.6, 6.6)
8	2.42 dd (12.6, 3.3)	2.99 dd (9.6, 5.1)
11 α	3.76 dd (14.4, 6.3)	3.11 dd (14.7, 3.0)
11 β	2.09 dd (14.4, 10.5)	2.02 dd (14.4, 12.3)
12	5.62 dd (10.5, 6.6)	5.44 dd (12.3, 2.4)
14	6.42 dd (1.2, 0.9)	6.42 dd (1.8, 0.9)
15	7.41 t (1.8)	7.41 t (1.8)
16	7.44 d (0.9)	7.49 d (0.9)
19	1.76 s	1.72 s
20	1.54 s	1.67 s
OH	7.07 s	6.92 s
CO ₂ CH ₃	3.86 s	3.85 s

Fig. 1. Key HMBC correlations of **4a** and **4b**.

related to H-6 α (δ 2.51), H-7 α (1.94), and H-20 (δ 1.54). It should be noted that there is no crossed peak between H-8 and H-12/H-19/H-20. Based on these data, the structure of **4a** was confirmed as deacetyl-1,10-didehydrosalvinorin G.

The coupling constants of H-6 and H-7 (Table 1) in **4a** are significantly different from those of **4b**. The *J* values

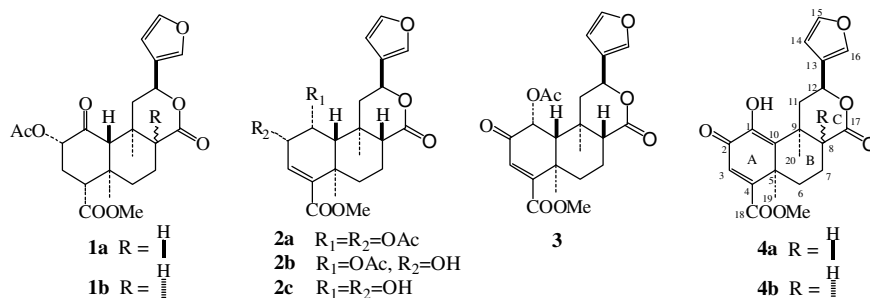
Table 2

¹³C NMR data (75 MHz, CDCl₃) for **4a**, **4b**, **6a**, **6b**, **7a**, **7b**, **8a**, **8b**, **9a**, and **9b** [δ (ppm)]

Carbon	4a	4b	6a ^a	6b ^b	7a ^b	7b ^b	8a ^b	8b ^b	9a ^c	9b ^c
1	145.3	145.1	208.9	209.2	201.9	202.3	203.7	204.1	202.2	202.6
2	180.9	180.7	74.4	74.4	74.9	75.1	75.9	76.2	75.2	75.5
3	127.6	128.2	34.5	33.8	30.6	30.5	31.8	31.5	28.3	28.1
4	159.0	157.5	53.1	52.3	53.4	52.6	50.7	50.2	59.7	59.1
5	43.3	42.3	42.6	42.6	42.0	42.1	41.9	42.0	42.5	42.6
6	32.0	28.3	38.1	34.2	38.1	33.8	38.0	33.8	38.0	33.8
7	18.1	21.9	18.1	17.5	18.1	17.5	18.1	17.6	17.9	17.4
8	50.6	44.8	51.3	45.2	51.3	45.2	51.4	45.3	51.2	45.1
9	39.4	37.6	35.3	34.5	35.4	34.7	35.2	34.6	35.4	34.6
10	138.5	140.0	63.8	63.6	63.9	63.9	64.4	64.5	63.6	63.5
11	42.4	36.8	43.5	48.2	43.3	47.9	43.3	48.0	43.2	47.9
12	71.5	70.8	71.9	70.0	72.1	70.1	72.1	70.1	72.0	70.1
13	126.0	124.4	125.3	123.5	125.1	123.3	125.2	123.3	125.1	123.2
14	108.6	108.4	108.3	108.4	108.4	108.5	108.4	108.5	108.3	108.5
15	143.8	143.7	143.8	143.6	143.7	143.6	143.7	143.6	143.8	143.6
16	139.4	139.6	139.3	139.6	139.4	139.7	139.4	139.7	139.4	139.7
17	171.6	173.2	171.0	173.4	171.3	173.7	171.4	173.7	170.9	173.5
18	166.0	165.4	171.8	172.1	175.8	176.2	61.6	61.6	200.6	200.9
19	31.3	30.3	16.5	15.3	16.4	15.3	16.6	15.5	17.9	16.7
20	18.8	24.4	15.2	24.6	15.2	24.6	15.3	24.8	15.1	24.4
CO ₂ CH ₃	52.7	52.6	51.9	51.6	—	—	—	—	—	—
–COCH ₃	—	—	—	—	170.0	169.9	170.1	169.9	170.0	169.8
–COCH ₃	—	—	—	—	20.5	20.5	20.6	20.6	20.6	20.6

^a Compound **6a** was isolated from the leaves of *Salvia divinorum*.⁷^b Compounds **6b**, **7a**, **7b**, **8a**, and **8b** were synthesized and reported by our group.^{15,16,21}^c The chemical shift assignments were based on the comparison with those in **1a** and **1b**.²⁷

of H-6 β (td, 13.5, 3.9) and H-7 α (dtd, 14.4, 3.3, 13.5) of **4a** indicate that both protons are axial. It has been reported that the protons in anti-periplanar relationships show stronger correlations in the COSY spectrum.³⁰ This was also evidenced by the protons (H-7 α and H-6 β , H-7 α and H-8) of **4a** in the COSY spectrum. On the other hand, only H-7 β and H-8 of **4b** exhibited stronger correlations in the COSY spectrum. These findings suggest that the B-ring in **4a** should be an identical chair conformation (Fig. 1), which is different from that of **4b**. Obviously, the double bond between C-1 and C-10 in **4a** and **4b** distorts the B-ring conformation to a different extent.



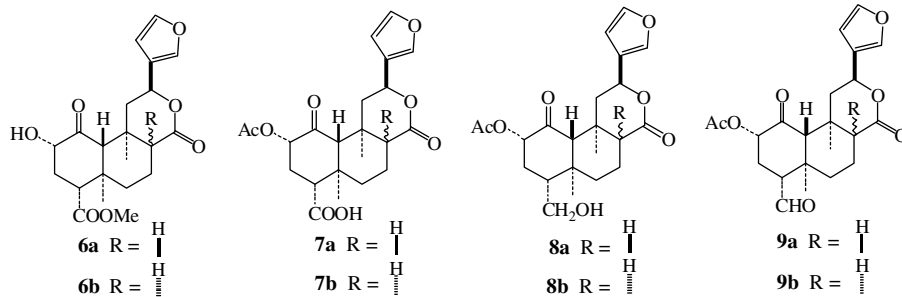
Compounds **4a** and **5** were screened for binding affinity at opioid receptors *in vitro*, as reported previously.⁷ Both compounds were inactive at mu, delta, and kappa opioid receptors at 3 μ M.

Salvinorin G (**3**) presents in *S. divinorum* in much lower level than **1a** and **2a**, and it showed a moderate binding

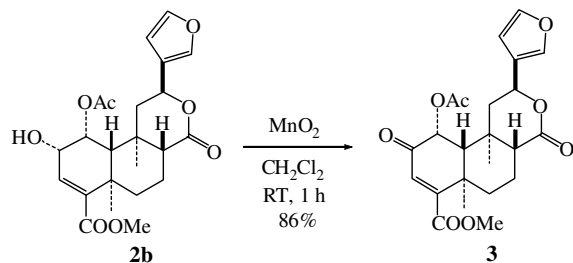
no epimerization occurs under the mild oxidation conditions as shown in Scheme 1.

Numerous salvinorin derivatives have been prepared in recent years for SAR study and improvement of KOR binding affinity.^{12–26} Compounds **6a**, **6b**, **7a**, **7b**, **8a**, and **8b** have served as key intermediates for C-2 and C-18 SAR studies. Among these compounds, only **6a** and **8a** were reported with full NMR assignments.^{30,13} However, **6a** was measured in acetone-*d*₆ at higher temperature (40 °C).³⁰ Furthermore, **1b** is the only compound with full ¹H and ¹³C NMR assignments in numerous 8-*epi*-salvinorin derivatives.¹³ Therefore, we assigned all ¹³C NMR

chemical shifts of **6a**, **6b**, **7a**, **7b**, **8a**, and **8b** in comparison with those of **1a** and **1b** (Table 2).²⁷ On the other hand, both aldehydes **9a** and **9b** were synthesized in our laboratory, and the incorrect ¹H and ¹³C NMR data of **9a** were presented in our previous Letter.²¹ The ¹³C NMR data of **9a** were revised and are shown in Table 2.



affinity at KOR.⁷ **3** can be prepared by oxidation of the natural occurring salvinorin D (**2b**)⁷ with manganese dioxide in an excellent yield (Scheme 2).³¹ This reaction not only confirms the structure of **3** but also demonstrates that



Scheme 2.

In conclusion, deacetyl-1,10-didehydrosalvinorin G (**4a**) was readily synthesized from salvinorin H (**2c**). The product obtained by the treatment of **1a** with hydroxides in MeOH¹⁸ has been unambiguously identified as 8-*epi*-deacetyl-1,10-didehydrosalvinorin G (**4b**). Finally, the conversion of salvinorin G (**3**) from salvinorin D (**2b**) provides an authentic sample with intact stereochemistry at C-8 for further confirmation of **4a**.

Acknowledgments

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Supplementary data

¹H and ¹³C NMR spectra of **4a**. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2008.01.065.

References and notes

1. Siebert, D. J. *J. Ethnopharmacol.* **1994**, *43*, 53–56.
2. Ortega, A.; Blount, J. F.; Manchand, P. S. *J. Chem. Soc., Perkin Trans. 1* **1982**, 2505–2508.
3. Valdes, L. J., III.; Butler, W. M.; Hatfield, G. M.; Paul, A. G.; Koreeda, M. *J. Org. Chem.* **1984**, *49*, 4716–4720.
4. Valdes, L. J., III.; Chang, H.-M.; Visger, D. C.; Koreeda, M. *Org. Lett.* **2001**, *3*, 3935–3937.
5. Munro, T. A.; Rizzacasa, M. A. *J. Nat. Prod.* **2003**, *66*, 703–705.
6. Bigham, A. K.; Munro, T. A.; Rizzacasa, M. A.; Robins-Browne, R. M. *J. Nat. Prod.* **2003**, *66*, 1242–1244.
7. Lee, D. Y. W.; Ma, Z.; Liu-Chen, L.-Y.; Wang, Y.; Chen, Y.; Carlezon, W. A.; Cohen, B. *Bioorg. Med. Chem.* **2005**, *13*, 5635–5639.
8. Harding, W. W.; Tidgewell, K.; Schmidt, M.; Shah, K.; Dersch, C. M.; Snyder, J.; Parrish, D.; Deschamps, J. R.; Rothman, R. B.; Prisinzano, T. E. *Org. Lett.* **2005**, *7*, 3017–3020.
9. Shirota, O.; Nagamatsu, K.; Sekita, S. *J. Nat. Prod.* **2006**, *69*, 1782–1786.
10. Roth, B. L.; Baner, K.; Westkaemper, R.; Siebert, D. J.; Rice, K. C.; Steinberg, S.; Ernsberger, P.; Rothman, R. B. *Proc. Natl. Acad. Sci.* **2002**, *99*, 11934–11939.
11. Sheffler, D. J.; Roth, B. L. *Trends Pharmacol. Sci.* **2003**, *24*, 107–109.
12. Chavkin, C.; Sub, S.; Jin, W.; Stewart, J.; Zjawiony, J. K.; Siebert, D. J.; Toth, B. A.; Hufeisen, S. J.; Roth, B. L. *J. Pharmacol. Exp. Ther.* **2004**, *308*, 1197–1203.
13. Munro, T. A.; Rizzacasa, M. A.; Roth, B. L.; Toth, B. A.; Yan, F. *J. Med. Chem.* **2005**, *48*, 345–348.
14. Beguin, C.; Richards, M. R.; Wang, Y.; Chen, Y.; Liu-Chen, L.-Y.; Ma, Z.; Lee, D. Y. W.; Carlezon, W. A.; Cohen, B. M. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 2761–2765.
15. Lee, D. Y. W.; Karnati, V. V. R.; He, M.; Liu-Chen, L.-Y.; Kondaveti, L.; Ma, Z.; Wang, Y.; Chen, Y.; Beguin, C.; Carlezon, W. A.; Cohen, B. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 3744–3747.
16. Lee, D. Y. W.; He, M.; Kondaveti, L.; Liu-Chen, L.-Y.; Ma, Z.; Wang, Y.; Chen, Y.; Li, J.-G.; Beguin, C.; Carlezon, W. A.; Cohen, B. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 4169–4173.
17. Harding, W. W.; Tidgewell, K.; Byrd, N.; Cobb, H.; Dersch, C. M.; Butelman, E. R.; Rothman, R. B.; Prisinzano, T. E. *J. Med. Chem.* **2005**, *48*, 4765–4771.
18. Munro, T. A.; Goetchi, G. W.; Roth, B. L.; Vortherms, T. A.; Rizzacasa, M. A. *J. Org. Chem.* **2005**, *70*, 10057–10061.
19. Harding, W. W.; Schmidt, M.; Tidgewell, K.; Kannan, P.; Holden, K. G.; Dersch, C. M.; Rothman, R. B.; Prisinzano, T. E. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 3170–3174.
20. Beguin, C.; Richards, M. R.; Li, J.-G.; Wang, Y.; Xu, W.; Liu-Chen, L.-Y.; Carlezon, W. A.; Cohen, B. M. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 4679–4685.
21. Lee, D. Y. W.; He, M.; Liu-Chen, L.-Y.; Wang, Y.; Li, J.-G.; Xu, W.; Ma, Z.; Carlezon, W. A.; Cohen, B. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 5498–5502.
22. Tidgewell, K.; Harding, W. W.; Lozama, A.; Cobb, H.; Shah, K.; Kannan, P.; Dersch, C. M.; Parrish, D.; Deschamps, J. R.; Rothman, R. B.; Prisinzano, T. E. *J. Nat. Prod.* **2006**, *69*, 914–918.
23. Munro, T. A.; Duncan, K. K.; Staples, R. J.; Xu, W.; Liu-Chen, L.-Y.; Beguin, C.; Carlezon, W. A.; Cohen, B. M. *Beilstein J. Org. Chem.* **2007**, *3* <http://bjoc.beilstein-journals.org/content/pdf/1860-5397-3-1.pdf>.
24. Bikbulatov, R. V.; Yan, F.; Roth, B. L.; Zjawiony, J. K. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 2229–2232.
25. Holden, K. G.; Kidgewell, K.; Marquam, A.; Rothman, R. B.; Navarro, H.; Prisinzano, T. E. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 6111–6115.
26. Bikbulatov, R. V.; Stewart, J.; Jin, W.; Yan, F.; Roth, B. L.; Ferreira, D.; Zjawiony, J. K. *Tetrahedron Lett.* **2008**, *49*, 937–940.
27. Ma, Z.; Lee, D. Y. W. *Tetrahedron Lett.* **2007**, *48*, 5461–5464.
28. *Synthesis of 4a*. To a solution of **2c** (12 mg, 31 μmol) in CH₂Cl₂ (3 ml) was added manganese dioxide (50 mg, 575 μmol), and the suspension was stirred at room temperature for 3 h. The solution was filtered and evaporated in vacuo. The residue was purified by silica gel column [CH₂Cl₂/AcOEt (10:1)] to give **4a** (6.2 mg, yield 52%) and **5** (2.5 mg, yield 21%). Data for **4a**: ¹H and ¹³C NMR data, see Tables 1 and 2; EI-MS *m/z* 386 (M⁺). Data for **5**: ¹H NMR (CDCl₃, 300 MHz) δ 7.46 (1H, br s, H-16), 7.43 (1H, br s, H-15), 6.41 (1H, br s, H-14), 6.38 (1H, s, H-3), 5.61 (1H, dd, *J* = 11.7 and 5.4 Hz, H-12), 4.33 (1H, d, *J* = 3.3 Hz, H-1), 3.84 (3H, s, COOCH₃), 2.54 (1H, br s, OH), 2.52 (1H, dd, *J* = 12.6 and 5.1 Hz, H-11a), 2.12–2.32 (3H, m, H-6a, H-7a, H-8), 1.70–1.94 (3H, m, H-7b, H-10, H-11b), 1.73 (3H, s, H-19), 1.53 (3H, s, H-20), 1.34 (1H, m, H-6b); ¹³C NMR (CDCl₃, 75 MHz) δ 198.9 (C-2), 171.4 (C-17), 166.4 (C-18), 162.3 (C-4), 143.9 (C-15), 139.4 (C-16), 127.1 (C-3), 125.4 (C-13), 108.3 (C-14), 71.8 (C-12), 70.1 (C-1), 54.0 (C-10), 52.5 (C-8), 52.0 (COOCH₃), 43.2 (C-11), 38.0 (C-5), 37.3 (C-9), 35.1 (C-6), 23.3 (C-19), 18.2 (C-7), 16.8 (C-20); EI-MS *m/z* 388 (M⁺).
29. Munro, T. A. The Chemistry of *Salvia divinorum*. Ph.D. Thesis, University of Melbourne, Victoria, Australia, 2006; p 212. [<http://eprints.infodiv.unimelb.edu.au/archive/00002327>].
30. Giner, J. L.; Kiemle, D. J.; Kutzreba, L.; Zjawiony, J. *Magn. Reson. Chem.* **2007**, *45*, 351–354.
31. The quaternary carbon at C-17 and the carbonyl carbon of the acetyl group in **3** should be reassigned as δ 170.9 and δ 169.7, respectively.